Synthesis of Asymmetrically Substituted Aminohalogenobenzimidazoles

Maria-Jose Camarasa, Paul L. Coe, A. Stanley Jones,* and Richard T. Walker Chemistry Department, University of Birmingham, Birmingham B15 2TT

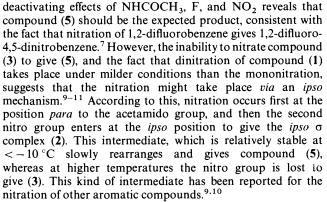
> 2-Fluoroacetanilide (1) upon treatment with $HNO_3-H_2SO_4$ at -5 to 0 °C gave 6-fluoro-4-nitroacetanilide (3) whereas at -20 to -10 °C the product was 6-fluoro-3,4-dinitroacetanilide (5). Although the formation of compound (5) could be accounted for by a conventional nitration mechanism, the fact that (3) could not be nitrated to give (5) and that (5) was formed at a lower temperature than was (3) suggested that the reaction might proceed *via* an *ipso* δ intermediate (2). Deacetylation of (5) followed by reduction and condensation with formic acid in the presence of HCl gave 6(5)-fluoro-5(6)formylaminobenzimidazole (11), 5(6)-amino-6(5)-fluorobenzimidazole (12) and 5-amino-4-chloro-6-fluorobenzimidazole (13). Reduction of (5) with tin followed by condensation with acetic acid gave 5(6)-amino-6(5)-fluoro-2-methylbenzimidazole (14). Concomitant reduction and cyclisation of 6chloro-2,4-dinitroaniline with formic acid and HCl gave a mixture of 7(4)-chloro-5(6)-formylaminobenzimidazole (15) and 5-amino-4,7-dichlorobenzimidazole (16).

Various benzimidazole derivatives have been used in the chemotherapy of viral diseases,¹ the most interesting being 5,6-dichloro-1- β -D-ribofuranosylbenzimidazole which inhibits RNA synthesis,^{2,3} influenza virus, vaccinia virus,⁴ and adenovirus.⁵ Benzimidazoles designed as guanine analogues have been shown to inhibit various guanine-sensitive test systems⁶ and thus inhibit virus multiplication.

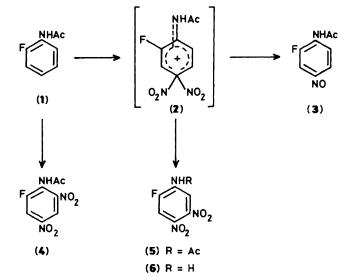
As part of a programme designed to elucidate structureactivity relationships amongst asymmetrically substituted aminohalogenobenzimidazoles, and nucleosides derived from them, we report here the syntheses of a series of substituted benzimidazoles.

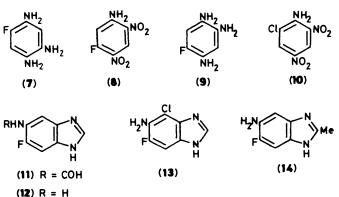
Acetylation of 2-fluoroaniline with acetic anhydride-pyridine gave 2-fluoroacetanilide (1) in quantitative yield. Nitration of compound (1) with fuming nitric acid-concentrated sulphuric acid at -20 to -10 °C gave the dinitro compound (5). On the other hand, when the reaction was carried out between -5 and 0 °C, only the mononitro derivative (3) was obtained. No further nitration of compound (3) to the dinitro compound (5) was observed and extended reaction times gave only decomposition to coloured compounds.

The formation of 6-fluoro-3,4-dinitroacetanilide (5) rather than 6-fluoro-2,4-dinitroacetanilide (4) is at first sight rather surprising. However, careful consideration of the activating/



Compound (5) was deacetylated with boiling 2Mhydrochloric acid to give 6-fluoro-3,4-dinitroaniline (6). Attempts to reduce this compound (6) to the triamino derivative (7) by a number of reagents such as tin(11) chloride,¹² catalytic hydrogenation, sodium dithionite,¹³ and hydrazine hydrate with palladium on charcoal,14 were unsuccessful owing to the instability of compound (7), which decomposed before it could be used in the cyclisation to the required benzimidazole. Subsequently, reduction and simultaneous closure of the imidazole ring was carried out by the Phillips procedure¹⁵ using tin, and hydrochloric acid with formic acid as the condensing agent, under a nitrogen or argon atmosphere. Under these conditions compound (6) gave 6(5)-fluoro-5(6)-formylaminobenzimidazole (11) and 5(6)amino-6(5)-fluorobenzimidazole (12) and, unexpectedly, 5-





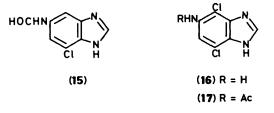
amino-4-chloro-6-fluorobenzimidazole (13), in 20, 46, and 13% yields respectively. Compound (13) was not produced when either benzimidazole (11) or (12) was treated with tin and hydrochloric acid and formic acid under the Phillips conditions.¹⁵ Thus, it seems that the entry of the chlorine atom in the benzimidazole ring takes place before the ring closure takes place. A similar reaction of compound (6) with acetic acid as the condensation reagent gave only 5-amino-6-fluoro-2-methylbenzimidazole (14) in 39% yield. The formation of compound (13) is unexplained, therefore; it is difficult to envisage electrophilic substitution of compound (5) by chlorine under the conditions of the reaction, and nucleophilic attack by chloride also appears unlikely.

Compound (11) was obtained as the only product (63% yield)when commercially available 5-fluoro-2,4-dinitroaniline (8) was reduced by catalytic hydrogenation in the presence of palladium on charcoal in formic acid, and then cyclised with formic acid. Attempts to obtain the 2-methylbenzimidazole derivative (14) by a similar procedure failed; the use of acetic acid in the catalytic hydrogenation led to the decomposition of the presumed intermediate triamine (9).

The assignment of structures to the benzimidazoles (11)-(14) was based upon analyses of hydrogen-fluorine coupling constants in the n.m.r. spectra (see Experimental section). The signals appearing at 7.48 p.p.m. in the spectrum of (11), at 7.20 p.p.m. for (12), at 7.32 p.p.m. for (13), and at 7.12 p.p.m. for (14), with H-F coupling constants of 11, 12, 10, and 12 Hz respectively, were assigned to the 7-H proton in the ortho position to the fluorine atom. The signals with an H-F coupling constant of 8 Hz which appeared at 8.32, 6.82, and 6.82 p.p.m. in compounds (11), (12), and (14), respectively, were assigned to 4-H. These H-F coupling constants are consistent with those corresponding to ortho and meta coupling constant values respectively.^{16.17} The presence of chlorine in compound (13) was deduced from its mass spectrum, and the position of the chlorine atom was assigned as 4 by comparison of the n.m.r. spectrum of compound (13) with that of (12), the former showing the disappearance of the signal at δ 6.82 p.p.m. corresponding to 4-H in (12).

Reduction of compound (8) with tin and hydrochloric acid in the presence of formic acid gave a mixture of compounds (11), (12), and (13) in yields of 11, 46, and 5% respectively. A similar reaction of compound (8), but with acetic acid instead of formic acid, gave the product (14) in 68% yield. These reactions confirmed the structures of compounds (11)—(14) and of the dinitro compound (5).

Other halogeno-substituted benzimidazoles were obtained by a similar synthetic route to that described above. Thus, when 6-chloro-2,4-dinitroaniline (10) was treated with tin and hydrochloric acid as reducing agent and formic acid as condensing agent, 7(4)-chloro-5(6)-formylaminobenzimidazole (15) and 5-amino-4,7-dichlorobenzimidazole (16) were obtained



in yields of 20 and 23% respectively. As in the case of the synthesis of the fluorobenzimidazoles already mentioned, a compound with an extra chlorine atom, namely (16), was isolated. However, catalytic hydrogenation of compound (10) in the presence of formic acid and subsequent condensation with formic acid gave compound (15) as the only isolatable reaction

product, in 19% yield. Acetylation of compound (16) gave the acetyl derivative (17) in 67% yield.

The structures of the chlorobenzimidazole derivatives (15)— (17) were established from their mass and n.m.r. spectra (see Experimental section). The signals appearing in the ¹H n.m.r. spectra at δ 7.51, 6.86, and 7.52 p.p.m. in compounds (15), (16), and (17), respectively, were assigned to 6-H. The disappearance of this signal in compound (17) indicated that the second chlorine group was located at position 4 of the benzimidazole ring.

The compounds described above have been tested for activity against herpes simplex virus types 1 and 2, vaccinia virus, vesicular stomatis virus, coxsackie virus B-4, and polio virus-1, but none showed any activity. Further tests of these compounds in other biological systems are in progress.

Experimental

Melting points were measured on an electrothermal m.p. apparatus or a Kofler block, and are uncorrected. N.m.r. spectra were measured at 100 MHz on a Varian XL100 spectrometer with $(CD_3)_2SO$ as solvent, mass spectra were obtained on a Kratos MS 80 RF mass spectrometer, and u.v. absorption spectra were recorded on a Perkin-Elmer 552 spectrophotometer. Analytical t.l.c. was carried out on silica gel 60 F_{254} (Merck), preparative layer chromatography on silica gel PF₂₅₄ (Merck), and column chromatography on Kieselgel 60, 70-230 mesh ASTM type 7734 (Merck).

6-Fluoro-4-nitroacetanilide (3).—To a mixture of fuming nitric acid (8.2 g) and concentrated sulphuric acid (24.5 g), stirred at -5 to 0 °C, 2-fluoroacetanilide (3.0 g) was added in portions over a period of 2 h. After being stirred for a further 6 h while the temperature was gradually raised to room temperature, the mixture was poured onto ice and allowed to stand at *ca*. 0 °C for 1 h. The resulting solid was then filtered off and washed successively with water, aqueous sodium hydrogen carbonate, and water to give the product (1.65 g, 43%) as a solid, m.p. 200—201 °C (from ethanol) (lit.,¹⁸ m.p. 203—204 °C); δ 2.20 (3 H, s, NCOCH₃), 8.02—8.26 (2 H, m, 3-H, 5-H), 8.46 (1 H, dd, J_{F,H} 8 Hz, 2-H, and 10.40 (1 H, br s, NH); *m/z* 198 (*M*⁺, 9%), 182(3), 156(75), 140(15), 126(20), 110(27), 98(3), 90(11), 83(17), 63(9), and 43(100).

6-Fluoro-3,4-dinitroacetanilide (5).—To a mixture of fuming nitric acid (13.5 g) and concentrated sulphuric acid (40.9 g) stirred at -20 to -10 °C, 2-fluoroacetanilide (5.0 g) was added in portions over a period of 3 h. The temperature was allowed to rise slowly to 0 °C and the mixture stirred at this temperature for 6 h. A work-up procedure similar to that described above afforded the *product* (5) (4.65 g, 59%) as yellow needles, m.p. 170—172 °C (from aqueous ethanol) (Found: C, 39.7; H, 2.3; N, 17.5. C₈H₆FN₃O₅ requires C, 39.51; H, 2.47; N, 17.28%); λ_{max} (ethanol) 302 (ε 7 600) and 239 nm (13 600); δ 2.20 (3 H, s, NCOCH₃), 8.28 (1 H, d, J_{F.H} 11 Hz, 5-H), 8.87 (1 H, d, J_{F.H} 7 Hz, 2-H), and 10.5 (1 H, br s, NH).

6-Fluoro-3,4-dinitroaniline (6).—Compound (5) (2.0 g) was added to 2M-hydrochloric acid (10 ml) and the mixture boiled under reflux for 4 h. After cooling the reaction mixture was neutralised with aqueous sodium hydrogen carbonate and cooled to 0 °C. The resulting solid was filtered off and crystallised from water to give the *product* (1.31 g, 83%) as needles, m.p. 135 °C (decomp.) (Found: C, 35.7; H, 2.0; N, 20.6. C₆H₄FN₃O₄ requires C, 35.82; H, 1.99; N, 20.89%); $\lambda_{max.}$ (ethanol) 372 (ε 9 800) and 243 nm (7 100); δ 7.13 (1 H, d, $J_{F,H}$ 8 Hz, 2-H), 7.22 (2 H, br s, NH₂), and 7.96 (1 H, d, $J_{F,H}$ 12 Hz, 5-H).

6(5)-Fluoro-5(6)-formylaminobenzimidazole (11), 5(6)-Amino-6(5)-fluorobenzimidazole (12), and 5-Amino-4-chloro-6-fluorobenzimidazole (13).—(a) To the dinitro derivative (6) (2.0 g) in a mixture of concentrated hydrochloric acid (40 ml) and formic acid (2.5 ml) at 70 °C under nitrogen, tin (6.21 g) was added in the form of strips over a period of 1.5 h. The temperature was raised to 110 °C and the reaction mixture boiled under reflux for 2 h while adding formic acid (5 ml). After cooling, propan-2-ol (50 ml) was added and the solution adjusted to pH 8-9 with aqueous ammonia. The precipitate of tin compounds was filtered off through Celite and the filtrate evaporated to dryness to give a dark solid which was purified by column chromatography using ethyl acetate-ethanol (5:1) as the eluant. From the faster-moving fractions, 5-amino-4-chloro-6fluorobenzimidazole (13) (0.24 g, 13%) was isolated as white needles by sublimation in vacuo, m.p. 218 °C (decomp.) (Found: C, 45.3; H, 3.0; N, 22.3. C₇H₅ClFN₃ requires C, 45.28; H, 2.69; N, 22.64%); λ_{max} (0.1M-NaOH) 293 (ϵ 3150) and 268 nm (3 080); $\lambda_{max}(0.1\text{m-HCl})$ 298 (ϵ 2 990) and 248 nm (2 620); δ 4.94 (2 H, s, NH₂), 7.32 (1 H, d, J_{F,H} 11 Hz, 7-H), and 8.02 (1 H, s, 2-H); m/z 187 (M^+ + 2, 30%), 185 (M^+ , 100), 152(3), 150(10), 123(19), 96(11), 76(5), 52(8), and 44(37).

The slower-moving fractions yielded a mixture of compounds (11) and (12) that was separated by column chromatography, using ethyl acetate-ethanol-acetic acid (4:0.8:1) as the eluant. From the resulting faster-moving fractions, 6(5)-fluoro-5(6)formylaminobenzimidazole (11) (0.36 g, 20%) was isolated as an amorphous solid, m.p. > 220 °C (Found: M^+ 179.0492. C₈H₆FN₃O requires *M*, 179.0495); λ_{max} (0.1M-NaOH) 277 (ε 9 613) and 269 nm (10 130); λ_{max} (0.1M-HCl) 285 (ε 5 540) and 283 nm (5 490); λ_{sh} 240 nm; δ 7.48 (1 H, d, $J_{F,H}$ 11 Hz, 7-H), 8.22 (1 H, s, 2-H), 8.32 (1 H, d, J_{F,H} 8 Hz, 4-H), 8.36 (1 H, s, HCON), 10.08 (1 H, br s, NH), and 12.4 (1 H, br s, NH); m/z 179 $(M^+, 70\%)$, 123(30), 104(7), 97(29), 76(12), 70(14), and 52(14). The slower-moving fractions afforded 5(6)-amino-6(5)-fluorobenzimidazole (12) (0.69 g, 46%) m.p. > 220 °C upon crystallisation from ethyl acetate-hexane (Found: M^+ 151.0549. $C_7H_6FN_3$ requires *M*, 151.0546); λ_{max} (0.1M-NaOH) 296 (ϵ 6 170) and 249 nm (3 100); λ_{max} (0.1M-HCl) 299 (ϵ 1 240), 277 (4 590), 272 (4 550), and 239 nm (3 530); & 5.75-6.6 (3 H, br, 5-NH₂, 1-NH), 6.86 (1 H, d, J_{F,H} 8 Hz, 4-H), 7.20 (1 H, d, J_{F,H} 12 Hz, 7-H), and 7.92 (1 H, s, 2-H); m/z 152 $(m^+ + 1, 10\%)$, 151 $(M^+, 100)$, 150(12), 124(11), 123(8), 104(5), 97(21), 75(10), 70(12), and 62(4).

(b) This procedure was similar to method (a) except that 5-fluoro-2,4-dinitroaniline (8) was used as the starting material and the reaction was carried out under argon. On work-up, the yields of the products isolated were: (11) (0.20 g, 11%), (12) (0.7 g, 46%), and (13) (0.1 g, 5.5%).

(c) The dinitro derivative (8) (2.5 g) was hydrogenated in a mixture of methanol (150 ml) and formic acid with palladiumcharcoal catalyst at ca. 20 °C at 3 atm. for 22 h. The catalyst was filtered off and the filtrate evaporated to dryness. To the residue was added formic acid (15 ml) and the solution boiled for 8 h. The solvent was removed by evaporation and the residue purified by column chromatography using ethyl acetateethanol (5:1) as the eluant. Collection of the appropriate fractions gave compound (11) (1.39 g, 63%), identical with that prepared by methods (a) and (b).

5(6)-Amino-6(5)-fluoro-2-methylbenzimidazole (14).—(a) To a mixture of concentrated hydrochloric acid (30 ml) and acetic acid (2 ml), was added compound (5) (2.0 g) and the mixture heated to 70 °C under nitrogen. Then tin (4.92 g) in the form of strips was added in portions over a period of 1.5 h, and the mixture then boiled under reflux for 2 h, cooled, diluted with water (20 ml) and propan-2-ol (62 ml), and adjusted to pH 9 with aqueous ammonia. Insoluble tin compounds were filtered off through Celite and the filtrate evaporated to dryness to give a dark solid. This was purified by column chromatography using ethyl acetate-methanol (10:1) as the eluant and finally by preparative t.l.c. using ethyl acetate-methanol (8:1) as the eluant. The *product* (14) was obtained as a white solid (0.51 g, 39%), m.p. 155—157 °C upon crystallisation from ethyl acetate-hexane (Found: M^+ , 165.0707. C₈H₈FN₃ requires M, 165.0702); λ_{max} (0.1M-NaOH) 295 (ε 5 560) and 246 nm (3 280); λ_{max} (0.1M-HCl) 294 (ε 700), 278 (4 960), 274 (5 140), 270 (4 830), and 233 nm (3 310); δ 2.42 (3 H, s, CH₃), 4.42—5.12 (3 H, br, NH₂, NH), 6.82 (1 H, d, J_{F,H} 8 Hz, 4-H), and 7.12 (1 H, d, J_{F,H} 12 Hz, 7-H); *m/z* 165 (M^+ , 100%), 164(80), 137(5), 123(8), 110(2), 97(16), 82(10), 70(8), and 60(40).

(b) This procedure was similar to method (a) except that compound (8) (2.0 g) was used as the starting material and the reaction mixture was heated under reflux for 3 h. On work-up the residue was purified by column chromatography using ethyl acetate-ethanol (5:1) as the eluant to give compound (14) (0.92 g, 68%), identical with that prepared by method (a).

7(4)-Chloro-5(6)-formylaminobenzimidazole (15) and 5-Amino-4,7-dichlorobenzimidazole (16).-(a) To a mixture of concentrated hydrochloric acid (92 ml) and formic acid (6 ml) was added 6-chloro-2,4-dinitroaniline (10) (5.0 g) and the mixture heated to 70 °C under argon. Tin (14.3 g) in the form of strips was added in portions over a period of 1.5 h and the mixture was then boiled under reflux for 6 h while adding formic acid (15 ml). On work-up as previously described, a dark solid was obtained. This was purified by column chromatography using ethyl acetate-methanol (10:1) as the eluant. From the faster-moving fractions 5-amino-4,7-dichlorobenzimidazole (16) (1.08 g, 23.5%), m.p. > 265 °C was obtained (Found: C, 41.9; H, 2.5; N, 30.1. C₇H₅Cl₂N₃ requires C, 41.58; H, 2.47; N, 29.79%); λ_{max} (0.1м-NaOH) 301 (ε 2 273) and 270 nm (3 520); λ_{max} (0.1M-HCl) 309 (ϵ 1 050) and 252 nm (3 530); δ 5.32 (2 H, br s, NH₂), 6.86 (1 H, s, 6-H), and 8.05 (1 H, s, 2-H); m/z 205 $(M^+ + 4, 7\%)$, 203 $(M^+ + 2, 44)$, 201 $(M^+, 100)$, 166(10), 139(19), 112(9), 103(18), 87(7), and 76(9).

The slower-moving fractions afforded 7(4)-chloro-5(6)formylaminobenzimidazole (15) (0.89 g, 20%) as an amorphous solid, m.p. > 220 °C (Found: C, 49.4; H, 3.0; N, 21.3. $C_8H_6CIN_3O$ requires C, 49.10; H, 3.06; N, 21.48%); δ 7.51 (1 H, d, 6-H), 8.00 (1 H, d, J 2 Hz, 4-H), 8.30 (1 H, s, 2-H), 8.36 (1 H, d, J 2 Hz, HCON, collapses to s on D₂O shake), and 10.39 (1 H, br s, NH); m/z 195 (M^+ - 1, 25%), 193(72), 179(9), 177(26), 169(34), 167(100), 130(38), 103(22), 88(11), and 76(20).

(b) The dinitro compound (10) (1.0 g) was hydrogenated in a mixture of methanol (150 ml) and formic acid (7 ml) with palladium-on-charcoal catalyst at 3 atm. at ca. 20 °C for 23 h. Following removal of the catalyst and the solvent, the residue thus obtained was dissolved in formic acid (20 ml) and the solution boiled under reflux for 5 h. Removal of the solvent and column chromatography of the residue using ethyl acetateethanol (5:1) as the eluant gave compound (15) (0.13 g, 15%), identical with that obtained in (a).

5-Acetylamino-4,7-dichlorobenzimidazole (17).—Compound (16) (0.64 g) and dimethylaminopyridine (0.05 g) was dissolved in dimethylformamide (10 ml) and acetic anhydride (20 ml) and the solution allowed to stand at *ca*. 20 °C for 2 days. The solvent was removed by evaporation and the residue crystallised from ethanol–ethyl acetate to give the *product* (17) (0.52 g, 67%) (Found: C, 44.6; H, 3.0; N, 17.5. C₉H₇Cl₂N₃O requires C, 44.26; H, 2.86; N, 17.21%); λ_{max} (0.1M-NaOH) 283 (ϵ 7 590) and 278 nm (8 150); λ_{max} (0.1M-HCl) 261 nm (8 970); δ 2.08 (3 H, s, NCOCH₃), 7.52 (1 H, s, 6-H), 8.38 (1 H, s, 2-H), and 9.64 (1 H, br s, NH); *m/z* 247 (*M*⁺ + 4, 3%), 245 (*M*⁺ + 2, 12), 243 (*M*⁺, 35), 210(27), 208(52), 205(15), 203(66), 201(100), 172(11), 166(12), 139(10), and 103(9).

Acknowledgement

We thank The Consejo Superior de Investigaciones Científicas of Spain for a research grant (to M. J. C.).

References

- 1 I. Tamm and L. A. Caliguiri in 'International Encyclopedia of Pharmacology and Therapeutics,' Section 61, Chemotherapy of Virus Diseases, Pergamon Press, Oxford 1972, vol. I, ch. 2.
- 2 E. Egyhazi, J. Mol. Biol., 1974, 84, 173.
- 3 P. B. Sehgal, J. E. Darnell, and I. Tamm, *Cell.*, 1976, 9, 473; P. B. Sehgal, E. Derman, G. R. Molloy, I. Tamm, and J. E. Darnell, *Science*, 1976, 194, 431.
- 4 I. Tamm and J. R. Overman, Virology, 1957, 3, 185.
- 5 I. Tamm, M. M. Nemes, and S. Osterhout, J. Exp. Med., 1960, 111, 339.
- 6 H. B. Gillespie, M. Engleman, and S. Graff, J. Am. Chem. Soc., 1954, 76, 3531.

- 7 G. C. Finger, R. H. Shiley, and D. R. Dickenson, *Illinois Geological* Survey Reports, 1978, 501, 93.
- 8 R. I. Thompson, J. Immunol., 1947, 55, 345; I. Tamm, K. Folkers, C. H. Shunk, and F. Horsfall, J. Exp. Med., 1954, 99, 227; D. A. J. Tyrrell and I. Tamm, J. Immunol., 1955, 75, 43.
- 9 C. L. Perrin and G. A. Skinner, J. Am. Chem. Soc., 1971, 93, 3389.
- 10 R. C. Hahn and D. L. Strack, J. Am. Chem. Soc., 1974, 96, 4337.
- 11 M. W. Galley and R. C. Hahn, J. Am. Chem. Soc., 1974, 96, 4339.
- 12 F. D. Bellamy and K. Ou, Tetrahedron Lett., 1984, 25(8), 839.
- 13 L. F. Fieser and M. Fieser, J. Am. Chem. Soc., 1934, 56, 1565.
- 14 B. M. Adger and R. G. Young, Tetrahedron Lett., 1984, 25(45), 5291.
- 15 J. Phillips, J. Chem. Soc., 1928, 2393.
- 16 E. Pretsch, T. Clerc, J. Seibl, and W. Simon in 'Tabellen zur Strukturaufklärung Organischer Verbindungen mit Spektroskopischen Methoden,' Springer Verlag, Berlin-Heidelberg-New York, 1976.
- 17 K. L. Kirk and L. A. Cohen, J. Org. Chem., 1969, 34, 384; Z. Kazimierczuk, L. Dudycz, R. Stolarski, and D. Shugar, J. Carbohydr. Nucleosides Nucleotides, 1981, 8(2), 101.
- 18 B. M. Wepster and P. E. Verkade, Recl. Trav. Chim., 1949, 68, 77.

Received 1st August 1986; Paper 6/1565